

DESCRIPTION

NIOSOME HAVING METAL PORPHYRIN COMPLEX EMBEDDED THEREIN,
PROCESS FOR PRODUCING THE SAME AND DRUG WITH THE USE THEREOF

TECHNICAL FIELD

The present invention relates to a niosome having a metalloporphyrin complex embedded therein and, more particularly, to a niosome having a metalloporphyrin complex embedded therein acting as an anticancer agent or an antioxidant in living bodies, and a process for producing the same.

BACKGROUND ART

Many active oxygen species generated in living bodies are said to participate in many diseases such as inflammation, nervous system diseases, arteriosclerosis, cancer, and diabetes. Usually, in order to maintain a balance, the living bodies produce radical scavenging enzymes such as superoxide dismutase (SOD), catalase, and the like to combat these active oxygen species.

However, cancer cells in living bodies are known to contain a large number of $O_2^{\cdot-}$ radicals. Therefore, enzyme activities are considered to be reduced in the cancer cells.

On the other hand, diseases such as inflammation, nervous system diseases, arteriosclerosis, and diabetes are thought to be caused by an increase in active species such as $O_2^{\cdot-}$ radicals due to imbalanced secretion of radical scavenging enzymes such as SOD, catalase, and the like.

Metalloporphyrin complexes are reported to exhibit high SOD activity. Therefore, if administered to a living body, a metalloporphyrin complex is expected to effectively scavenge active oxygen species such as $O_2^{\cdot-}$ radicals and protect the living

body from hindrance brought about by the active oxygen species.

However, since administering a metalloporphyrin complex independently to a living body has many problems from the viewpoint of safety and effects, no metalloporphyrin complexes have been used as a drug.

The present invention has been completed in view of these problems and has an object of providing a means for safely administering a metalloporphyrin complex to a living body and causing the SOD activity possessed by metalloporphyrin complex to be effectively exhibited.

A further object of the present invention is to provide an anticancer agent that can replace anticancer agents such as clinically used cisplatin and mitomycin C causing serious side effects by selectively exhibiting an effect on cancer cells and also to provide an antioxidant for treating diseases other than cancer such as inflammation, nervous system diseases, arteriosclerosis, and diabetes in which active oxygen species are considered to be involved.

DISCLOSURE OF THE INVENTION

As a result of extensive studies in order to develop a method for decreasing the concentration of $O_2^{\cdot -}$ radicals in cancer cells using the SOD activity of metalloporphyrin complexes, the present inventors have found that the metalloporphyrin complexes can be safely administered while maintaining superior SOD activity and can be retained in the blood by being embedded in a niosome. This finding has led to completion of the present invention.

Specifically, the present invention provides a niosome having a metalloporphyrin complex-embedded therein comprising a cationized metalloporphyrin complex and a niosome-forming substance.

The present invention further provides a process for producing a niosome having a metalloporphyrin complex embedded therein comprising mixing a cationized

metalloporphyrin complex and a niosome-forming substance and treating the mixture with supersonic waves in a medium.

The present invention further provides a process for producing a metalloporphyrin complex-embedding niosome comprising reacting a cationized metalloporphyrin complex and an anionic surfactant to produce an ion complex, mixing the resulting ion complex with a niosome-forming substance, and treating the mixture with supersonic waves in a medium.

The present invention also provides a drug comprising the metalloporphyrin complex-embedding niosome as an effective component.

BRIEF DESCRIPTION OF THE DRAWING

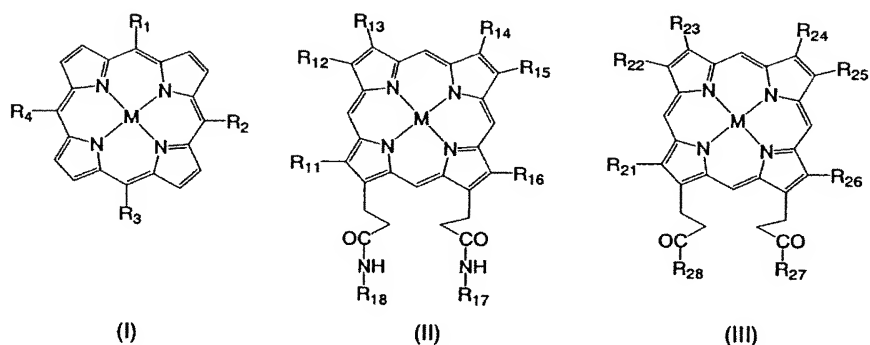
Figure 1 shows results of tests for confirming an anticancerous effect using FeT2MPyP as a metalloporphyrin complex and Pluronic F-88 as a niosome-forming substance, wherein the solid line connecting black triangle marks indicates the result of ion complex 2 (FeT2MPyP/4SAS), the dotted line connecting black square marks indicates the result of Pluronic F-88, solid line connecting black square marks indicates the result of CDDP, the solid line connecting black diamond marks indicates the result of MMC, and the solid line connecting black circle marks indicates the result of the invention product 9.

BEST MODE FOR CARRYING OUT THE INVENTION

In the present invention, “metalloporphyrin complex-embedding niosome” indicates a niosome in which a metalloporphyrin complex is incorporated, formed by a niosome-forming substance such as a mixture of a nonionic surfactant, a cholesterol, and the like, in which either a part of the metalloporphyrin complex may be present outside the niosome membrane or the entire amount of the metalloporphyrin complex is embedded in the niosome membrane.

The metalloporphyrin complex-embedding niosome of the present invention comprises a cationized metalloporphyrin complex and a niosome-forming substance.

The cationized metalloporphyrin complex of the metalloporphyrin complex-embedding niosome (hereinafter referred to from time to time as “Por-embedding niosome”) of the present invention is a porphyrin complex having a group containing a cationic nitrogen atom as a substituent and includes, for example, the following compounds (I), (II), and (III),



wherein at least one of R_1 to R_4 is a group selected from an N-lower-alkylpyridyl group, a lower-alkylammoniophenyl group, and an N-lower-alkylimidazolyl group, R_{11} to R_{16} and R_{21} to R_{26} are lower alkyl or lower alkoxy groups, R_{17} and R_{18} are groups selected from an N-lower-alkylpyridyl group, a lower-alkylammoniophenyl group, and an N-lower-alkylimidazolyl group, and R_{27} and R_{28} indicate N-alkylammonio groups.

Compounds having a methylpyridyl group for R_1 in the above formula (I) such as 5,10,15,20-tetrakis(2-methylpyridyl)porphyrin (T2MPyP), 5,10,15,20-tetrakis(4-methylpyridyl)porphyrin (T4MPyP), 5,10,15,20-tetrakis(3-methylpyridyl)porphyrin (T3MPyP), and the like, compounds having a methylpyridylamideethyl group for R_2 in the above formula (II) such as [1,3,5,8-tetramethyl-2,4-divinyl-6,7-di(4-methylpyridylamideethyl)]porphyrin

(PPIX-DMPyAm) and the like can be given as more specific examples.

As the metal coordinating to these cationized porphyrin complexes, iron (Fe), manganese (Mn), cobalt (Co), copper (Cu), molybdenum (Mo), chromium (Cr), iridium (Ir), and the like are preferable.

Among the above-mentioned metalloporphyrin complexes, the cationized porphyrin complex coordinated with a metal shown by the formula (I) can be synthesized according to a method described in K. Kalyanasundaram, *Inorg. Chem.*, 23, 2453 (1984), A. D. Adler et al., *J. Inorg. Nucl. Chem.*, 32, 2443 (1970), T. Yonetani et al., *J. Biol. Chem.*, 245, 2988 (1970), P. Hambright, *Inorg. Chem.*, 15, 2314 (1976), and the like.

The cationized porphyrin complex shown by the formula (II) coordinated with a metal can be synthesized according to a method described in E. Tsuchida, H. Nishide, H. Yokoyama, R. Young, and C. K. Chang, *Chem. Lett.*, 1984, 991, and the like.

The Por-embedding niosome of the present invention can be produced by using only the above-mentioned cationized porphyrin complexes, but more preferably is produced by using an ion complex obtained by reacting a cationized metalloporphyrin complex with an anionic surfactant (hereinafter referred to simply as “ion complex”).

As examples of the anionic surfactant used for producing the ion complex, alkali metal salts of a fatty acid and alkali metal salts of alkyl sulfuric acid are preferable. As specific examples, alkali metal salts of a fatty acid such as lauric acid (LAS), myristic acid (MAS), palmitic acid (PAS), stearic acid (SAS), and oleic acid (OAS), and alkali metal salts of an alkyl sulfuric acid such as dodecylsulfuric acid (SDS), tetradecylsulfuric acid (STS), hexadecylsulfuric acid (SHS), octadecylsulfuric acid (SOS), and the like can be given. Sodium, potassium, and the like are preferable as the metal components of the alkali metal salts of a fatty acid and alkyl sulfuric acid.

In order to form the above ion complex, a cationized metalloporphyrin complex is mixed with an anionic surfactant in a suitable solvent at a molar ratio of the

cationized metalloporphyrin complex to the anionic surfactant of from about 1:1 to 1:20.

On the other hand, the above cationized metalloporphyrin complex or the ion complex made from the cationized metalloporphyrin complex and the anionic surfactant is mixed with a niosome-forming substance to produce a Por-embedding niosome by a conventional method of producing a niosome.

As the niosome-forming substance, many compounds reported to produce a niosome can be used. Nonionic surfactants and mixtures of a nonionic surfactant and a cholesterol or triacylglycerol are preferably used.

As the nonionic surfactant, polyoxyethylene sorbitan fatty acid esters, sorbitan fatty acid esters, polyoxyethylene polyoxypropylene copolymers, and the like can be used either individually or in combination. As specific examples of the nonionic surfactant, Tween-61 (TW61), Tween-80, and Span 80 (manufactured by NOF Corp.), Pluronic F-88 (F88) (manufactured by ADEKA Corp.), and the like can be given.

As examples of the cholesterol, cholesterol, α -cholestanol, β -cholestanol, cholestane, desmosterol (5,24-cholestadiene-3 β -ol), sodium cholate, and cholecalciferol can be given.

In order to produce a Por-embedding niosome from the above cationized metalloporphyrin complex or the ion complex and the niosome-forming substance, these components must first be added to a suitable solvent and sufficiently mixed.

In forming a niosome, the cationized metalloporphyrin complexes shown by the formulas (II) or (III) can be embedded in the niosome as is, but the cationized metalloporphyrin complex shown by the formulas (I) is preferably embedded into a niosome in the form of an ion complex.

The amount of the cationized metalloporphyrin complex and the niosome-forming substance used in forming the niosome, in terms of the amount (mol) of the niosome-forming substance for one mol of the cationized metalloporphyrin

complex or the ion complex, is preferably from 10 to 500 mols, and particularly preferably from 50 to 300 mols.

The niosome can be formed by a commonly known process, for example, a process comprising dissolving and mixing the above components in a volatile solvent, removing only the volatile solvent by vaporization, dissolving the residue in an appropriate aqueous solvent such as purified water or a physiological saline solution, and vigorously stirring or supersonically treating the solution to form a Por-embedding niosome.

If necessary, a solution of a pharmaceutically effective component or a certain type of culture medium may be used instead of the aqueous solvent to produce a Por-embedding niosome in which such a pharmaceutically effective component or a culture medium is incorporated.

As a result of structural analysis of the Por-embedding niosome obtained in this manner using a fluorescence spectrum, dynamic light scattering measurement, and the like, as described later, it was confirmed that a cationized metalloporphyrin complex portions are present on the surface of the niosome or in hydrophilic areas of lipids and the like and alkyl group side chains of the surfactant are embedded in hydrophobic areas of lipids.

It was also confirmed that the niosome has a particle size of 100 nm or less, and particularly can be made into endoplasmic reticula with a diameter of 30 nm or less, which is a size enabling the niosome to reach cells when incorporated into the body.

In addition, as a result of the test for confirming the anticancerous effect of the Por-embedding niosome as described later, the Por-embedding niosome was confirmed to be more effective than a simple cationized metalloporphyrin complex used as a raw material of the niosome. The effect was confirmed to be higher than the effect of cisplatin and mitomycin C which are presently used as anticancer agents.

Moreover, the evaluation of the SOD activity of the Por-embedding niosome

confirmed that the Por-embedding niosome exhibits an SOD activity equivalent to the simple cationized metalloporphyrin complex used as a raw material of the niosome, indicating usability as an SOD compound possessing superior retainability in the blood.

As mentioned above, the Por-embedding niosome of the present invention has excellent anticancer activity and can be used as an anticancer agent in the field of cancer therapy.

Therefore, it is possible to treat cancer of a patient by administering the Por-embedding niosome to the patient by direct dosage, intravenous administration, subcutaneous administration, and the like.

The Por-embedding niosome of the present invention also possesses a superior effect as an antioxidant and can be used for protecting living bodies from disorders brought about by active oxygen species such as, for example, inflammation, nervous system disorders, arteriosclerosis, and diabetes.

Therefore, if the Por-embedding niosome is administered to a patient suffering from inflammation, nervous system disorders, arteriosclerosis, or diabetes by direct dosage, intravenous administration, subcutaneous administration, or the like means, that patient as cancer can be treated.

EXAMPLE

The present invention will be described in more detail by way of Examples and Test Examples which should not be construed as limiting the present invention.

Example 1

Synthesis of iron[5,10,15,20-tetrakis(2-methylpyridyl)porphyrin] (FeT2MPyP)

(1) After heating 500 ml of propionic acid to 110°C with stirring, 15 ml (0.158 mol) of 2-pyridylcarboxyaldehyde was added. Then, 12 ml (0.173 mol) of pyrrole was slowly added dropwise using an injector and the mixture was refluxed at 110°C for one

hour to effect a cyclization-condensation reaction. After the reaction, the reaction solution was allowed to cool to room temperature and the solvent was evaporated. The residue was neutralized, washed, and purified by column chromatography (alumina basic type-I, chloroform) to obtain 5,10,15,20-tetrakis(2-pyridyl)porphyrin (amount: 1.1 g, yield: 4.4%).

$^1\text{H-NMR}$ $\delta\text{H}(\text{CDCl}_3, \text{ppm})$: -2.82 (H of pyrrole NH, 2H), 7.72-9.14 (H of pyridine, 16H), 8.87 (H of pyrrole, 8H)

UV-vis λ_{max} (chloroform, nm): 418, 513, 544, 586, 645

FAB-Mass(m/z): 619, 620

(2) Next, 0.2 g (3.2×10^{-4} mol) of 5,10,15,20-tetrakis(2-pyridyl)porphyrin obtained in (1) above was added to and dissolved in 150 ml of dimethylformamide under an argon (Ar) atmosphere. After the further addition of iron bromide (FeBr_2) produced from 0.2 g of iron and 5 ml of 48% hydrobromic acid, the mixture was refluxed for four hours. After the reaction, the reaction solution was allowed to cool to room temperature and the solvent was evaporated.

The residue was purified by fractionation using a water-chloroform mixture, followed by column chromatography (alumina basic type-I, methanol) to obtain iron[5,10,15,20-tetrakis(2-pyridyl)porphyrin] (amount: 0.21 g, yield: 94%) which is a precursor of the target compound.

UV-vis λ_{max} (methanol, nm): 408, 512, 566

FAB-Mass(m/z): 672

(3) 0.1 g of iron[5,10,15,20-tetrakis(2-pyridyl)porphyrin] obtained in (2) above and 6 ml of methyl p-toluenesulfonate were added to 30 ml of dimethylformamide and the mixture was refluxed at 130°C for five hours. After refluxing, the reaction solution was allowed to cool to room temperature and the solvent was evaporated. The residue

was purified by extraction and column chromatography (alumina basic type-I, methanol) to obtain the target compound FeT2MPyP (yield: 91%).

UV-vis λ_{max} (water, nm): 408, 584

Elemental analysis (%): Found: C75.11, H3.97, N17.66, C/N4.25

Calcd: C77.58, H4.24, N18.12, C/N4.28

(4) Iron[5,10,15,20-tetrakis(4-methylpyridyl)porphyrin] (FeMT4MPyP) was obtained in the same manner as in the steps (1) to (3) above except for using 4-pyridylcarboxyaldehyde instead of 2-pyridylcarboxyaldehyde in the step (1). Other metalloporphyrin complexes of [5,10,15,20-tetrakis(4-methylpyridyl)porphyrin] (MT4MPyP) and [5,10,15,20-tetrakis(2-methylpyridyl)porphyrin] (MT2MPyP) can be synthesized in the same manner as above.

Example 2

Synthesis of manganese[5,10,15,20-tetrakis(2-methylpyridyl)porphyrin] (MnT2MPyP)

(1) After heating 500 ml of propionic acid to 110°C, 15 ml of 2-pyridylcarboxyaldehyde was added, following which 12 ml of pyrrole was slowly added. After the addition of all the amounts, the mixture was refluxed for one hour. After the reaction, the reaction mixture was allowed to cool, evaporated, neutralized, and washed. The residue was purified by column chromatography (basic alumina, chloroform) and the like to obtain purple crystals of 5,10,15,20-tetrakis(2-pyridyl)porphyrin (amount: 1.68 g, yield: 7.08%).

$^1\text{H-NMR}$ $\delta\text{H}(\text{CDCl}_3, \text{ppm})$: -2.8 (H of pyrrole NH, 2H), 7.7 (H of pyridine (4 position), 4H), 8.1 (H of pyridine (5 position), 4H), 8.2 (H of pyridine (6 position), 4H), 9.2 (H of pyridine (4 position), 4H), 8.8 (H of pyrrole, 8H)

UV-vis λ_{max} (chloroform, nm): 418, 513, 544, 586, 645

FAB-Mass(m/z): 619, 620

(2) 1.98 g of manganese acetate tetrahydrate was added to 200 ml of a dimethylformamide (DMF) solution containing 500 mg of 5,10,15,20-tetrakis(2-pyridyl)porphyrin obtained in (1) after argon replacement, and the mixture was refluxed for three hours in an argon stream. After the reaction, the reaction mixture was allowed to cool and subjected to evaporation, fractionation with water/chloroform, and drying under reduced pressure to obtain manganese[5,10,15,20-tetrakis(2-pyridyl)porphyrin] (amount: 409 mg, yield: 75.2%).

UV-vis λ_{max} (chloroform, nm): 461, 555

FAB-Mass(m/z): 672

(3) Next, 200 mg of the above manganese[5,10,15,20-tetrakis(4-pyridyl)porphyrin] was reacted with 12 ml of methyl p-toluenesulfonate at 130°C for eight hours. After the reaction, the reaction mixture was allowed to cool and subjected to fractionation with water/chloroform and column chromatography ((1) acidic alumina and (2) basic alumina, methanol) to obtain the target compound MnT2MPyP (amount: 153 mg).

UV-vis λ_{max} (water, nm): 456, 555

(4) Manganese[5,10,15,20-tetrakis(4-methylpyridyl)porphyrin] (MnT4MPyP) was obtained in the same manner as in the steps (1) to (3) above except for using 4-pyridylcarboxyaldehyde instead of 2-pyridylcarboxyaldehyde in the step (1). Other metalloporphyrin complexes of [5,10,15,20-tetrakis(2-methylpyridyl)porphyrin] (MT2MPyP) and [5,10,15,20-tetrakis(4-methylpyridyl)porphyrin] (MT4MPyP) can be synthesized in the same manner as above.

Example 3

Synthesis of Por-embedding niosome (1)

(1) 1.4 mg (1 μmol) of FeT2MPyP, a cationized metalloporphyrin complex prepared in Example 1, and 0.3 mg (1 μmol) of SAS, a surfactant, were put into a test tube and 10 ml of methanol was added as a solvent and mixed to obtain an ion complex 1 (FeT2MPyP/1SAS).

An ion complex 2 (FeT2MPyP/4SAS) and ion complex 3 (FeT2MPyP/4OAS) were also prepared in the same manner using 1.2 mg (4 μmol) of SAS and 1.2 mg (4 μmol) of OAS respectively instead of 1 μmol of SAS.

In the same manner, ion complexes 4 to 9 shown in Table 1 were prepared using MnT2MPyP, MnT4MPyP, FeT2MPyP + MnT2MPyP (0.5 μmol : 0.5 μmol), and FeT4MPyP + MnT4MPyP (0.5 μmol : 0.5 μmol) as cationized metalloporphyrin complexes and SAS or OAS as surfactants.

Table 1

Ion complex	Cationized metalloporphyrin	Surfactant	Cationized metalloporphyrin/Surfactant
1	FeT2MPyP [1 μmol]	SAS [1 μmol]	1/1
2	FeT2MPyP [1 μmol]	SAS [4 μmol]	1/4
3	FeT2MPyP [1 μmol]	OAS [4 μmol]	1/4
4	MnT4MPyP [1 μmol]	SAS [1 μmol]	1/1
5	MnT4MPyP [1 μmol]	SAS [4 μmol]	1/4
6	MnT2MPyP [1 μmol]	SAS [1 μmol]	1/1
7	MnT2MPyP [1 μmol]	SAS [4 μmol]	1/4
8	FeT2MPyP + MnT2MPyP (1:1) [1 μmol]	SAS [4 μmol]	1/4
9	FeT4MPyP + MnT4MPyP (1:1) [1 μmol]	SAS [4 μmol]	1/4

(2) Next, the above ion complex 1 (1 μmol), 0.131 g (100 μmol) of Tween-61, a niosome-forming substance, and 0.0386 g (100 μmol) of cholesterol were put into a test tube, to which a small amount of chloroform as a solvent was added and mixed. After

evaporation of the solvent to form a thin film, 10 ml of a physiological saline solution was added and the mixture was supersonically treated in an ice cooled bath under an argon atmosphere for 3 minutes using a probe-type supersonic-wave irradiation apparatus at 30 W. After the supersonic treatment, the product was allowed to stand for 30 minutes at 60°C and sterilized by filtration using a 0.22 µm diam. filter to obtain a Por-embedding niosome (FeT2MPyP/1SAS-embedding TW61-chol niosome; Invention Product 1).

(3) Another Por-embedding niosome (FeT2MPyP/4SAS-embedding TW61-chol niosome; Invention Product 2) was obtained in the same manner as in (2) above using the ion complex 2 (1 µmol). Por-embedding niosomes (Invention Products 3 to 8) shown in Table 2 can be obtained in the same manner using the ion complexes 1, 2, and 4 to 6.

Table 2

Ribosome having porphyrin complex embedded therein	Ion complex	Niosome-producing substance
FeT2MPyP/1SAS-embedding TW61-chol niosome (Invention Product 1)	1	Tween-61 and cholesterol (1:1)
FeT2MPyP/4SAS-embedding TW61-chol niosome (Invention Product 2)	2	Tween-61 and cholesterol (1:1)
FeT4MPyP/1SAS-embedding TW61-chol niosome (Invention Product 3)	1	Tween-61 and cholesterol (1:1)
FeT4MPyP/4SAS-embedding TW61-chol niosome (Invention Product 4)	2	Tween-61 and cholesterol (1:1)
MnT2MPyP/1SAS-embedding TW61-chol niosome (Invention Product 5)	6	Tween-61 and cholesterol (1:1)
MnT2MPyP/4SAS-embedding TW61-chol niosome (Invention Product 6)	7	Tween-61 and cholesterol (1:1)
MnT4MPyP/1SAS-embedding TW61-chol niosome (Invention Product 7)	4	Tween-61 and cholesterol (1:1)
MnT4MPyP/4SAS-embedding TW61-chol niosome (Invention Product 8)	5	Tween-61 and cholesterol (1:1)

Example 4

Synthesis of Por-embedding niosome (2)

The ion complex 2 (0.5 μmol) shown in Table 1, 0.54 g (50 μmol) of Pluronic F-88, a niosome-forming substance, and 0.0154 g (40 μmol) of cholesterol were put into a test tube, to which a small amount of chloroform (about 3 ml) as a solvent was added and mixed. After evaporation of the solvent to form a thin film, 10 ml of a physiological saline solution was added and the mixture was supersonically treated in an ice cooled bath for 30 minutes using a probe-type supersonic-wave irradiation apparatus at 30 W. After the supersonic treatment, the product was allowed to stand for 30 minutes at 50°C and sterilized by filtration using a 0.22 μm diam. filter to obtain a Por-embedding niosome (FeT2MPyP/4SAS-embedding F88-chol niosome; Invention Product 9).

Example 5

Measurement of Por-embedding niosome dynamic light scattering (1)

Dynamic light scattering of Por-embedding niosome was measured using a particle sizing system (“Nicomp 370”) in order to determine the particle size and particle size distribution of the Por-embedding niosome. The results of measuring dynamic light scattering of FeT2MPyP/1SAS-embedding TW61-chol niosome (Invention Product 1) and FeT2MPyP/4SAS-embedding TW61-chol niosome (Invention Product 2) synthesized according to Example 3 are shown in Table 3.

Table 3

Niosome having porphyrin complex embedded therein		First distribution peak		Second distribution peak	
		Particle size (nm)	Distribution ratio (%)	Particle size (nm)	Distribution ratio (%)
FeT2MPyP/1SAS-embedding TW61-chol niosome (Invention Product 1)	Number distribution	28.4	90	145	10
FeT2MPyP/4SAS-embedding TW61-chol niosome (Invention Product 2)	Number distribution	28.5	90	146	10

As a result, it can be seen that the number distribution 90% of particles had an average particle size of 28.5 nm and 10% had an average particle size of 146 nm. The results coincided with the results of the TEM inspection. It can also be seen from Table 4 that all Por-embedding niosomes had an average particle size of 100 nm or less, which, when administered, is small enough to pass through microvascular endothelium and reach the target cells.

Example 6

Measurement of Por-embedding niosome dynamic light scattering (2)

Dynamic light scattering of MnT2MPyP/1SAS-embedding TW61-chol niosome (Invention Product 5) was measured in the same manner as in Example 6. As a result, the niosome was found to have an average particle size of 28.5 nm (the distribution ratio: 90%, the balance of about 10% had an average particle size of 146 nm). The average particle size of MnT2MPyP/4SAS-embedding TW61-chol niosome (Invention Product 6) was 28.5 nm (the distribution ratio: 90%, the balance of about 10% had an average particle size of 146 nm). Both niosomes had an average particle size of 100 nm or less which is small enough to cause no problems when administered to a living body.

Example 7

Fluorescence spectrum measurement of Por-embedding niosome

(1) Fluorescence spectrum measurement of Por-embedding niosome was conducted using an “RF-5300PC” (manufactured by Shimadzu Corp.) in order to determine the place at which the embedded porphyrin complex is present in the niosome.

In the measurement, since a metalloporphyrin complex extinguishes fluorescence in general, cationized metal-free porphyrin complexes (hereinafter referred to as “metal-free complex”) were synthesized according to the methods of Examples 1, 2, etc. and used instead of the metalloporphyrin complexes as fluorescence probes. Metal-free complex-embedding niosomes were synthesized according to Example 3. Hereinafter, as abbreviation of a metal-free complex, H₂ is used instead of M in the formula of the cationized metalloporphyrin complex. For instance, the metal-free complex of metal[5,10,15,20-tetrakis(2-methylpyridyl)porphyrin] (MT2MPyP) is indicated as H₂T2MPyP, and the metal-free complex of metal[5,10,15,20-tetrakis(4-methylpyridyl)porphyrin] (MT4MPyP) is indicated as

H₂T4MPyP.

A fluorescence spectrum of cationized metal-free complexes-embedding niosome prepared in the above-described manner, for example, a fluorescence spectrum of H₂T2MPyP of various solutions containing H₂T2MPyP/4SAS-embedding TW61-chol niosome and the H₂T2MPyP/4SAS was measured at a wavelength in a range from 500 to 800 nm. As a result, a fluorescence spectrum having a peak at 645 nm was obtained using an aqueous solution containing H₂T2MPyP/4SAS-embedding TW61-chol niosome.

This fluorescence peak wavelength was the middle of the fluorescence wavelengths of H₂T2MPyP in water and methanol (fluorescence peak wavelength (short wavelength side) water < embedding niosome solution < methanol (long wavelength side)).

The fluorescence spectra of the H₂T2MPyP in various solutions such as methanol (peak: 642 nm), ethanol (peak: 643 nm), 2-propanol (peak: 645 nm), n-butanol (peak: 645 nm), ethylene glycol (peak: 642 nm) had the same waveform as above. However, the fluorescence spectra of H₂T2MPyP in water has a broad peak, exhibited a significant decrease in the strength, and shifted to a short wavelength side (630 nm). Based on these results, the H₂T2MPyP embedded in niosome is believed to be present in polar conditions resembling the conditions of the above alcohols, specifically, near the hydrophilic part of bimolecular membrane of which the conditions are slightly non-polar (slightly more hydrophobic) than the conditions of water. The result of H₂T2MPyP/1SAS-embedding TW61-chol niosome was also the same.

(2) Next, fluorescence spectra were measured using 8-anilino-1-naphthalenesulfonic acid (ANS), which is a fluorescence probe being present near the hydrophilic part of the niosome bimolecular membrane and acting as an indicator for the polarity, fluidity, and the like of the bimolecular membrane. All

fluorescence spectra of ANS in methanol, methanol/chloroform, and a solution of ANS-embedding TW61 niosome (no porphyrin complex is embedded) were similar to each other having a peak at 485nm (excitation wavelength: 385 nm). Next, a fluorescence spectrum of ANS was measured in a solution of TW61 niosome in which a porphyrin complex and ANS are embedded together. The spectrum did not have a peak at 485 nm, but showed two peaks at 450 nm and 500 nm, each having a peak fluorescence strength weaker than the peak fluorescence strength at 485 nm. The presence of Soret band absorption peaks of porphyrin complex in this region is believed to be the result of interaction between ANS and the porphyrin complex, which indicates the presence of the porphyrin complex near the ANS. Based on these results, the H₂T2MPyP embedded in niosome is believed to be present in polar conditions resembling the conditions of the above alcohols, specifically, near the hydrophilic part of the bimolecular membrane.

In a niosome in which the porphyrin complex is embedded together with 1,6-diphenyl-1,3,5-hexatriene (DPH), which is a fluorescent probe existing in the inside of a bimolecular membrane (hydrophobic part), no peak reduction was seen in the Soret band of porphyrin complex and in DPH peaks at 410 nm, 434 nm, and 457 nm, indicating that there was no interaction between DPH and the porphyrin complex and that the porphyrin complex did not exist near the DPH. Based on these results, the H₂T2MPyP embedded in niosome is believed to be present in polar conditions resembling the conditions of the above alcohols, specifically, near the hydrophilic part of the bimolecular membrane.

Example 8

Fluorescence depolarization measurement of Por-embedding niosome

Fluorescence depolarization of a metalloporphyrin complex-embedding niosome was measured using 1,6-diphenyl-1,3,5-hexatriene (DPH, 1 μ M) existing near the

hydrophilic part of a bimolecular membrane as a fluorescent probe (instrument: polarization auxiliary analyzer for an RF-5300PC and RF-540/5000 manufactured by Shimadzu Corp., excitation wavelength: 361 nm, fluorescence wavelength: 428 nm) at 5-60°C. In the relation of the fluorescence depolarization measurement temperature and the degree of polarization (a reverse sigmoidal curve) of the TW61-chol niosome containing DPH (blank), for example, a reduction of the fluorescence polarization degree was seen near the phase transition temperature ($T_c = 36^\circ\text{C}$) of the TW61-chol niosome bimolecular membrane. In the relation of the fluorescence depolarization measurement temperature and the degree of polarization of the FeT2MPyP/1SAS-embedding TW61-chol niosome containing DPH, although a reduction of the fluorescence polarization degree was seen near the phase transition temperature ($T_c = 37^\circ\text{C}$) in the same manner as in the blank, the polarization degree was large in a gel state. This is due to the interaction among FeT2MPyP/1SAS, Tween-61, and cholesterol, indicating that FeT2MPyP/1SAS is present in the niosome bimolecular membrane formed by Tween-61 and cholesterol.

The relation of the fluorescence depolarization measurement temperature and the degree of polarization of the MnT2MPyP/1SAS-embedding niosome containing DPH was seen to slightly shift to a high temperature side as compared with the blank and the polarization degree along the vertical axis increased in a gel state. These results are based on the interaction among MnT2MPyP/1SAS, Tween-61, and cholesterol and support the presence of MnT2MPyP/1SAS in the niosome bimolecular membrane formed by Tween-61 and cholesterol.

Example 9

Test of anticancer properties of Por-embedding niosome

The anticancer properties of the Por-embedding niosome of the present invention were examined by a cytotoxic test (cell-death test) according to the Alamar

Blue method .

Various Por-embedding niosomes (metalloporphyrin complex concentrations: 0, 12.5, 25, 50, and 100 μM) were used as the test samples. As reference samples, metalloporphyrin complexes (concentrations: 0, 12.5, 25, 50, and 100 μM) and niosomes (concentrations: 2,500, 5,000, 10,000, and 20,000 μM) which are respectively components of the Por-embedding niosomes were used. Both concentrations correspond to the Por-embedding niosome concentrations.

As comparative samples, currently used anticancer agents such as cisplatin (CDDP, concentration: 0, 10, 20, 40, 80 μM), mitomycin C (MCC, concentration: 0, 7.5, 15, 30, 60 μM), and the like were used with mouse lung cancer cells (Lewis Lung Carcinoma (LLC) supplied by RIKEN Gene Bank).

A specific test method was as follows. Cells were cultured in a DMEM medium to which 10% FBS was added, followed by counting of the number of cells and adjusting the cell concentration to obtain a cell suspension liquid (100 μl /well, number of cells: 1×10^4 cells/well). The cell suspension liquid was added to wells in a 96 well plate and the cells were cultured in a CO_2 incubator (5% CO_2) for 24 hours. After removing the medium from the well plate, previously prepared test sample solutions with a different concentration (active component concentration: 0 to 100 μM) were added in an amount of 100 μl /well, followed by incubation for 24 hours in a CO_2 incubator. After the addition of an alamar blue solution (10 μl /well) sterilized by filtration, the mixture was incubated for a further five hours. Then, absorbance (at a wavelength of 570 nm and a reference wavelength of 600 nm) was measured using a microplate reader.

The results obtained by using Pluronic F-88 niosome (Invention Product 9) in which FeT2MPyP was embedded as a Por-embedding niosome are shown in Figure 1. Based on the results, it can be seen that the LLC cell survival rate decreases as the test solution concentration increases, indicating effective anticancer properties. In

particular, the LLC cell survival rate when an ion complex of FeT2MPyP/4SAS was used at an FeT2MPyP concentration of 25 μ M or more was slightly lower than the rate achieved by CDDP or MMC. In addition, the LLC cell survival rate decreased as the concentration of the FeT2MPyP/4SAS-embedding F88-chol niosome added increased, specifically to as low as 10% or less at an addition concentration of 50 μ M, indicating anticancer properties superior to those of the currently-used CDDP and MMC.

The FeT2MPyP/1SAS-embedding TW61-chol niosome (Invention Product 1) and FeT2MPyP/4SAS-embedding TW61-chol niosome (Invention Product 2), both of which are Por-embedding niosomes, also showed anticancer properties superior to those of the currently-used CDDP and MMC. When the concentration was 50 μ M, for example, the anticancer effect increased in the order of currently-used anticancer agent < ion complex < cationized metalloporphyrin complex < metalloporphyrin complex-embedding niosome. The metalloporphyrin complex-embedding niosome can thus be concluded to be an excellent anticancer agent.

Example 10

Interaction between metalloporphyrin complex and hydrogen peroxide (H_2O_2)

A metalloporphyrin complex with a low molecular weight is reported to be easily decomposed in the presence of a large excess amount of hydrogen peroxide (H_2O_2) due to a high probability of porphyrin rings to interact with H_2O_2 because the porphyrin rings are exposed (R. F. Pasternack and B. Halliwell, J. Am. Chem. Soc., 101, 1026 (1979)). For this reason, the interaction of a low-molecular weight MnT2MPyP and a niosome (high-molecular weight) MnT2MPyP/1SAS-embedding TW61-chol niosome (Invention Product 5) and MnT2MPyP/4SAS-embedding TW61-chol niosome (Invention Product 6) with H_2O_2 was examined by UV-vis measurement. Specifically, these niosomes were interacted with H_2O_2 at different concentrations to measure a decay curve of the porphyrin complex Soret band absorption peak (456 nm) based on

the decomposition due to the interaction. The reaction rate constant ($k_{H_2O_2}$) was calculated from the decay curve. The interaction was then evaluated based on the half-life ($t_{1/2}$) calculated from the reaction rate constant. The results obtained by using copper/zinc superoxide dismutase (Cu/Zn-SOD) are also shown for reference. In the experiment, the samples were prepared according to the method described in Example 2, in which H_2O_2 concentrations of 1.2, 2.0, 3.0, and 30 mM, and a metalloporphyrin complex concentration of 5 μ M were used. The results are shown in Table 4.

Table 4

Metalloporphyrin complex system	$t_{1/2}$ (sec)
MnT2MPyP	730
MnT2MPyP/1SAS-embedding TW61-chol niosome (Invention Product 5)	1,005
MnT2MPyP/4SAS-embedding TW61-chol niosome (Invention Product 6)	806
Cu/Zn-SOD (comparative substance)	10

The results show that the $t_{1/2}$ increased in the order of Cu/Zn-SOD < MnT2MPyP < MnT2MPyP/4SAS-embedding TW61-chol niosome < MnT2MPyP/1SAS-embedding TW61-chol niosome, indicating that the MnT2MPyP having exposed porphyrin rings due to the low molecular weight has a large probability of interacting with H_2O_2 and is easily decomposed, whereas MnTM2PyP-embedding TW61-chol niosomes having a higher molecular weight are decomposed only with difficulty because porphyrin rings are not exposed due to the high molecular weight resulting in a low probability of interacting with H_2O_2 . The results coincided with the results of the fluorescence spectrum measurement in Example 7. In addition, it was shown that the $t_{1/2}$ of the MnT2MPyP/1SAS-embedding TW61-chol niosome is greater than the $t_{1/2}$ of Cu/Zn-SOD, indicating that the MnT2MPyP/1SAS-embedding TW61-chol niosome has greater H_2O_2 resistance than Cu/Zn-SOD.

Example 11

Evaluation of SOD activity of Por-embedding niosome (1)

The SOD activity (i.e. $O_2^{\cdot-}$ extinction activity) of Por-embedding niosome was evaluated according to the cytochrome c method ((1) J. M. Mccord and I. Fridovich, J. Biol. Chem., 244, 6049 (1969) and (2) J. Butler, W. H. Kopenol, E. Margoliash, J. Biol. Chem., 257, 10747 (1982)). A specific procedure was as follows: Five or more Por-embedding niosome solutions (A solutions) with different metalloporphyrin concentrations ranging from 0 to 1,000 μ M were prepared. A mixed solution (B solution) was prepared by mixing a 0.3 mM xanthine aqueous solution, a 60 μ M cytochrome c aqueous solution, and a 30 mM phosphoric acid buffer solution (7.8 pH), each in an amount of 20 ml, and adding 24 ml of purified water. 0.3 ml of A solution and 0.2 ml of purified water were added to 2.1 ml of B solution and the resulting mixture was allowed to stand at 25°C for 10 minutes. Then, 0.1 ml of a 7 μ g/ml catalase aqueous solution and 5 μ l of a 25 U/ml xanthine oxidase (XOD) aqueous solution were rapidly added to and mixed with the above mixture after standing to measure UV-vis at 550 nm (an absorption peak based on ferrocycytochrome c formation) over time. The ultimate concentrations of the components in the test solutions were 0 to 100 μ M of the metalloporphyrin complex, 0.05 mM of xanthine, 2.5 U/ml of XOD, 10 μ M of cytochrome c, and 0.23 μ g/ml of catalase. The UV-vis was also measured for a solution without addition of Por-embedding niosome (blank) in the same manner.

The ferrocycytochrome c formation rates in a solution not containing Por-embedding niosome (vo) and a solution containing Por-embedding niosome (vi) were determined from the relationship between the time and the absorbance at 550 nm in the UV-vis measurement over time. The inhibition coefficient (IC) was calculated according to the following formula to determine the concentration of metalloporphyrin when IC = 50% (IC_{50}) from the relationship between the concentration of

metalloporphyrin complex and IC. The resulting IC₅₀ value was regarded as the SOD activity. The smaller the IC₅₀ value, the higher the SOD activity. The SOD activity of MnT4MPyP and the like was evaluated in the same manner for reference.

$$\text{Inhibition coefficient (IC)} = 1 - (v_i/v_o)$$

v_o: rate of formation of ferrocytochrome c in a solution not containing Por-embedding niosome.

v_i: rate of formation of ferrocytochrome c in a solution containing Por-embedding niosome

Table 5 shows IC₅₀ values for various metalloporphyrin complex systems. In the Table, IC₅₀ value of MnT4MPyP is a value disclosed by Fridovich et al. (I. Batinic-Haberle, L. Benov, and I. Fridovich, J. Biol. Chem., 273, 24251 (1998).

Table 5

Metalloporphyrin complex system	IC ₅₀ (μM)
MnT2MPyP/1SAS-embedding TW61-chol niosome (Invention Product 5)	0.32
MnT2MPyP/4SAS-embedding TW61-chol niosome (Invention Product 6)	0.18
MnT2MPyP	0.18
MnT4MPyP	0.43
MnT4MPyP	0.7 ^{*1}

*1: I. Batinic-Haberle, L. Benov, and I. Fridovich, J. Biol. Chem., 273, 24251 (1998)

The results show that IC₅₀ values of MnT2MPyP/1SAS-embedding TW61-chol niosome and MnT2MPyP/4SAS-embedding TW61-chol niosome are close to reported values of low molecular weight MnT2MPyP, indicating that they exhibit an effective

SOD activity.

Example 12

Evaluation of SOD activity of Por-embedding niosome (2)

The SOD activity (i.e. $O_2^{\cdot-}$ extinction activity) of Por-embedding niosome was evaluated according to the stopped flow method of Riley et al. (D. P. Riley, W. L. Rivers, and R. H. Weiss, *Anal. Biochem.*, 196, 344 (1991)). A specific procedure was as follows: 60 mM HEPES/HEPES Na buffer solutions (8.1 pH) containing Por-embedding niosome at different concentrations were rapidly mixed with a dimethyl sulfoxide solution of potassium superoxide, which is an $O_2^{\cdot-}$ generation source, at 36°C to measure over time an attenuation of absorbance at 245 nm due to $O_2^{\cdot-}$ (a decay curve of $O_2^{\cdot-}$ extinction reaction). The relationship of \ln (absorbance) and time was determined from the decay curve. The apparent rate constant was calculated from the inclination of the resulting relationship. The rate constant (k_{cat}) of the $O_2^{\cdot-}$ extinction reaction was then determined from the inclination of the relationship of the metalloporphyrin complex concentration and the apparent rate constant.

Table 6 shows k_{cat} values for various metalloporphyrin complex systems. In the Table, the k_{cat} value of MnT4MPyP is a value disclosed by Ohse, Kawakami, et al. (T. Ohse, S. Nagaoka, Y. Arakawa, H. Kawakami, and K. Nakamura, *J. Inorg. Biochem.*, 85, 201 (2001)).

Table 6

Metalloporphyrin complex system	k_{cat} ($\text{M}^{-1}\text{s}^{-1}$)
MnT2MPyP/1SAS-embedding TW61-chol niosome (Invention Product 5)	3.1×10^7
MnT2MPyP/4SAS-embedding TW61-chol niosome (Invention Product 6)	2.0×10^7
MnT2MPyP	2.1×10^7
MnT4MPyP	1.4×10^7
MnT4MPyP	2.2×10^7 *2

*2: T. Ohse, S. Nagaoka, Y. Arakawa, H. Kawakami, and K. Nakamura, J. Inorg. Biochem., 85, 201 (2001)

The results show that k_{cat} values of MnT2MPyP/1SAS-embedding TW61-chol niosome and MnT2MPyP/4SAS-embedding TW61-chol niosome are close to reported values of low molecular weight MnT2MPyP, indicating that they exhibit an effective SOD activity.

INDUSTRIAL APPLICABILITY

The Por-embedding niosome of the present invention can react with superoxide anionic radicals ($\text{O}_2^{\cdot-}$) and can reduce these radicals without fail.

Therefore, the Por-embedding niosome can exhibit an excellent effect of treating cancer by reducing $\text{O}_2^{\cdot-}$ in cancer cells. In addition, since the effect is selective, the Por-embedding niosome can be used as a novel anticancer agent without side effects.

The Por-embedding niosome of the present invention also possesses an SOD activity and can be retained in the blood while exhibiting a superior antioxidation effect. The Por-embedding niosome can thus protect living bodies from hindrance brought about by active oxygen species.